

## S-Trap™ Micro Ultra-High Recovery Protocol

- 1) Elute protein from IPs or dissolve protein in 25  $\mu$ L of high recovery urea-SDS lysis/solubilization buffer: 5% SDS, 8 M urea, 100 mM glycine\* pH 7.55. TEAB can also be used.\*
- 2) Clarify sample as needed by centrifugation for 8 min at 13,000 g.
- 3) Reduce and alkylate disulfides. Avoid high temperatures are not recommended due to urea.
- 4) Add 2.5  $\mu$ L 12% phosphoric acid to the lysate solubilized in 25  $\mu$ L SDS buffer.
- 5) Add 165  $\mu$ L of S-Trap binding buffer (90% MeOH, 100 mM final TEAB, pH 7.1) into the S-Trap micro column. It will not flow through.
- 6) **The next two steps must be done as quickly as possible.** Add 1 or 2  $\mu$ g of trypsin to the acidified SDS and immediately mix by pipetting up and down, then immediately transfer the acidified lysate plus trypsin into the S-Trap binding buffer within the spin column. Again mix by pipetting up and down.
- 7) Spin in bench-top centrifuge in a standard 1.7 mL sample tube at 4,000 g until all solution has passed through. Remove flow through.
- 8) Wash by adding 150  $\mu$ L S-Trap buffer to the spin column and centrifuging through. Remove flow through. Repeat three times. Protein will not be lost during washes.
- 9) Add 0.5  $\mu$ g of trypsin in 25  $\mu$ L of 50 mM TEAB, pH 8 to the top of the protein trap. The protein trapping matrix is highly hydrophilic and will absorb the solution. However, ensure there is no bubble atop the protein trap.
- 10) Cap the spin column loosely and incubate in a clean tube for 1 hr at 47 °C for trypsin. Most preferably use a water bath or thermomixer. **DO NOT SHAKE. The cap MUST NOT form an air-tight seal.**
- 11) Elute peptides with 40  $\mu$ L each of 50 mM TEAB and then 0.2% aqueous formic acid. Add the first TEAB elution to the trypsin solution prior to any centrifugation. Centrifuge elutions through at 4,000 g.
- 12) Elute hydrophobic peptides with 35  $\mu$ L 50% acetonitrile, 0.2% formic acid.
- 13) Dry down peptides and resuspend as desired (buffer A or MALDI matrix).

\*Glycine will be removed during washing and is preferable to TEAB to limit carbamylation from urea.



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